ORIGINAL PAPER

Complexation of Dimethyl 2,3-Naphthalenedicarboxylate with 2-hydroxypropyl- α -, - β - and - γ -cyclodextrins in Aqueous Solution by Fluorescence, Circular Dichroism and Molecular Mechanics

Ruben Usero · Carolina Alvariza · María José González-Álvarez · Francisco Mendicuti

Received: 30 January 2008 / Accepted: 27 February 2008 / Published online: 4 June 2008 © Springer Science + Business Media, LLC 2008

Abstract Fluorescence, circular dichroism and molecular mechanics have been used to study the complexation of 2.3dimethyl naphthalenedicarboxylate with 2-hydroxypropyl- α -, - β and - γ -cyclodextrins (*HP*CDs) in aqueous solution. Emission spectra upon excitation of the naphthalenedicarboxylate group show two bands whose intensity ratio R is quite sensitive to polarity. From the change of R and lifetimes averages $\langle \tau \rangle$ with HPCD concentration and temperature were obtained the stoichiometry, the association constants and the enthalpy and entropy changes during the complexation. R, $<\tau>$ and the fluorescence anisotropies (r) extrapolated at [HPCD] $\rightarrow \infty$ allows us to estimate the polarity and microviscosity of the media surrounding the guest when complexed. In addition, the analysis of quenching and induced circular dichroism experiments and molecular mechanics calculations in the presence of water, provide information about the forces responsible for the complexation and the geometry of the complexes.

Keywords Cyclodextrins · Inclusion complex · Fluorescence · Circular dichroism · 2,3-Napthalenedicarboxylate · 2,3-Dimethyl naphthoate · Molecular mechanics

R. Usero · C. Alvariza · M. J. González-Álvarez ·
F. Mendicuti (⊠)
Departamento de Química Física, Universidad de Alcalá, 28871 Alcalá de Henares,
Madrid, Spain
e-mail: francisco.mendicuti@uah.es

Introduction

Cyclodextrins (CDs) are donut-shaped molecules capable of forming inclusion complexes with low molecular weight compounds and polymer guests [1-4]. CD cavities are relatively hydrophobic and the exterior surfaces are quite hydrophilic. Both microviscosity and polarity of the medium surrounding the guest molecule are substantially modified upon complexation. These changes frequently modify the spectroscopic characteristics of chromophore containing guests. Fluorescence emission intensity [5-13], excimer formation [14-18], fluorescence anisotropy [12, 19–22], the relative intensity of some bands of the emission spectra, [20-27] fluorescence decay [5, 8, 9, 12, 13, 21, 22, 27-29], energy transfer [30-32], fluorescence quenching [11-13, 20, 21, 30, 33], etc. may be affected upon inclusion. The analysis of the changes in these properties provides information about the structure of the complexes formed and allows us to determine quantitatively the stoichiometry, formation constants and thermodynamic parameters accompanying the association.

It is well-known that α -, β -, and γ HPCD are chiral hosts that do not absorb in the UV–Vis region. However, an achiral chromophore guest may exhibit an induced circular dichroism (ICD) when forming a guest:CD complex [34]. In fact, ICD has also been used to determine the stability constants and stoichiometry for such complexes. In addition, ICD provides information about the geometry of the complex, as the sign and strength of the ICD signal are related to the distance and relative orientation of the guest with respect to the CD framework [35].

Molecular modeling [36], (molecular mechanics [10, 20, 27, 30, 37–42] and/or molecular dynamics [41–48]) has

also been extensively used as an additional tool for clarifying the inclusion mechanism as well as to extending the knowledge on the driving forces involved in such processes [1, 2].

Steady-state and time resolved fluorescence techniques combined with molecular mechanics calculations were used in a previous study on the complexation of 2,3-naphthalenedicarboxylate (23DMN) with natural α -, β - and γ CDs [49]. Spectra for 23DMN in the absence or in the presence of CDs exhibit two electronic bands whose ratio is very sensitive to the medium polarity. The association constants and the ΔH^0 and ΔS^0 were obtained from the change of this ratio with [CD] and temperature. All the complexes showed 1:1 stoichiometry. Binding constants at 25 °C for 23DMN complexes with α -, β - and γ CDs were 20±6, $1,060\pm25$ and 40 ± 10 M⁻¹ respectively. Complexation with α - and β CDs exhibited $\Delta H^0 < 0$; the inclusion of 23DMN into γ CD showed $\Delta H^0 > 0$. Whereas the 23DMN: α CD complex formation was accompanied by a $\Delta S^0 < 0$, the complexes with β - and γ CD were achieved with a $\Delta S^0 > 0$. The $\Delta S^0 < 0$ was attributed to the partial 23DMN penetration into the α CD; $\Delta S^0 > 0$ pointed towards a deep penetration of the guest into the β - and γ CD cavities. Values of polarity surrounding 23DMN in the complexes estimated from the values of R extrapolated at $[CD] \rightarrow \infty$, also agree with the latter statements. 23DMN, in the complexes with α -, β - and γ CDs, is surrounded by media with effective dielectric constants (ε) of 70, 49 and 58 respectively. ε for 23DMN/ β CD was similar to that expected for the inner BCD obtained previously with other similar probes; the one for 23DMN/yCD implied that the γCD cavity is more polar [20-22, 25, 26]. However, the large value obtained for 23DMN with α CD, whose cavity is well-known to be more hydrophobic, showed that a large portion of 23DMN may be exposed to the water. These results may agree with the entropy changes, the extrapolated fluorescence anisotropy values and they might not disagree with the quenching experiments [49]. Molecular mechanics calculations also demonstrated that a strong repulsive energetic barrier impedes 23DMN from entering inside the α CD cavity.

In a similar way the complexation of 2-methyl carboxylate (2MN) with 2-hydroxypropyl- α -, - β - and - γ -cyclodextrins was studied and compared with that of their natural counterparts α -, β - and γ CDs [20, 21, 50]. Emission spectra of 2MN also exhibited two typical bands. Results showed identical stoichiometry (1:1) for the three 2MN complexes with α -, β - and γ HPCDs. Complexes with α - and β HPCDs resulted more stable than those formed with their counterparts α - and β CDs. Complexes with γ HPCD and γ CD showed similar stability at 25 °C [20, 21]. Complexation of 2MN with α HPCD is enthalpy driven, with β HPCD both the entropy and enthalpy terms favor the process, whereas the formation of the complex with $\gamma HPCD$ is entropically governed. Complexation processes of 2MN with natural CDs are accompanied by larger negative enthalpy changes than the ones obtained with their hydroxypropylated counterparts. The entropic terms accompanying the complexation, are more favorable (β - and γ *HP*CDs) or less unfavorable (α *HP*CDs) than with their natural counterparts. The differences in thermodynamics parameters upon complexation with respect to the natural CDs should mainly be caused by the differences in cavity sizes, which are apparently larger for the substituted CDs. Results revealed effective dielectric constants of the inner HPCD cavities of 51, 44 and 56 for α -, β - y γ *HP*CD respectively. These values differ from the α -, β -, and γ CDs which behave more polar as the number of glucopyranose units increases [20]. Fluorescence anisotropy, quencher lifetimes, average lifetimes and molecular modeling also agreed with a total penetration of 2MN inside the HPCD cavities.

In this work we investigate the complexation of 23DMN with 2-hydroxypropyl- α -, - β and - γ -cyclodextrins (HPCDs) by using several steady-state and time-resolved fluorescence techniques, circular dichroism, as well as molecular mechanics calculations (MM). Stoichiometries, binding constants and ΔH^0 and ΔS^0 , as well as information about the location of the guest in the complex, the polarity and microviscosity of the medium surrounding it, intermolecular interactions, etc. were obtained. Molecular mechanics methods were also used to support and strengthen most of the experimental evidence.

Materials and methods

Reagents and solutions

23DMN guest (Aldrich, 99%) and α -, β - and γ HPCDs (Aldrich, 0.6 substitution degrees) were used as received. Karl-Fisher analysis for HPCDs revealed water contents by mass of approx. 7.0, 7.1 and 6.6% respectively. Deionized water (Milli-Q) and other solvents, n-alcohols $H(CH_2)_nOH$ with n=1-6 (Aldrich spectrophotometric grade or $\geq 98\%$) were checked for impurities by fluorescence before use. Water 23DMN/HPCD solutions were prepared by weight from a doubly filtered (cellulose filters, Millipore, 1 $\mu m \emptyset$) guest saturated solution $([23DMN] = \sim 10^{-7} M)$. [HPCD] ranged from 0 to $3.724 \times$ 10^{-2} , 1.792×10^{-2} and 3.187×10^{-2} M for α -, β - and yHPCD respectively. The contents of the cells were stirred for ~48 h prior measuring. For induced circular dichroism measurements, in order to avoid signal saturation at the wavelength of the maximum of the intense ¹B_b band (~245 nm), the stock 23DMN guest solution was diluted approximately fifteen times.

Apparatus

Steady-state fluorescence measurements were recorded on an SLM 8100 Aminco Spectrofluorimeter equipped with a Xe lamp, a double (single) concave grating monochromator at the excitation (emission) path and Glan–Thompson prism polarizers in both paths. The photomultiplier was cooled by a Peltier system. The excitation and emission slit widths were 8 nm. The fluorescence decay measurements were achieved on a time correlated single photon counting (TCSPC) FL900 Edinburgh Instruments Spectrometer with a thyratron-gated lamp filled with H₂. Data acquisition was carried out by using a multichannel time detector and a time window width of 125 ns with a total of 10,000 counts at the intensity maximum. The instrumental response function was regularly obtained by measuring the scattering of a Ludox solution.

Induced circular dichroism (ICD) spectra were obtained by using a JASCO J-715 spectropolarimeter. Recorded spectra were the average of 2 scans taken at the speed of 5 nm/min with an 8 s time response. The bandwidth was set at 2 nm and the sensitivity and resolution were fixed at 20 mdeg and 0.2 nm respectively. All measurements were performed at 25 °C in 10 mm length cells.

Determination of association constants

Association constants for a 1:n guest:host complex (G: $HPCD_n$), whose global equilibrium can be written as

$$n HPCD + 23DMN \rightleftharpoons 23DMN : HPCD_n$$
 (1)

were determined by steady-state fluorescence measurements from the change of parameter R with [*HPCD*], by fitting the experimental data to the following equations [49]:

$$R = \frac{R_0 + R_\infty \Phi K [HPCD]_0^n}{1 + \Phi K [HPCD]_0^n}$$
(2)

and

$$\frac{1}{R - R_0} = \frac{1}{K\Phi(R_0 - R_\infty)} \frac{1}{[HPCD]_0^n} + \frac{1}{R_0 - R_\infty}$$
(3)

where R_0 and R_∞ are the values of R for the free and complexed guest. R in both equations is defined as the I $(\lambda_2)/I(\lambda_1)$ ratio, by assuming that the emission spectra show bands whose intensity maxima are centered at λ_1 and λ_2 . Thus $\Phi = I_\infty(\lambda_1)/I_0(\lambda_1)$, where $I_0(\lambda_1)$ and $I_\infty(\lambda_1)$ are the emission intensities (at λ_1) for the guest at [*HP*CD]=0 and extrapolated at [*HP*CD] $\rightarrow \infty$. From the change with [HPCD] of the weighted average lifetime, defined as

$$\langle \tau \rangle = \frac{\sum_{i=1}^{n} A_{i} \tau_{i}^{2}}{\sum_{i=1}^{n} A_{i} \tau_{i}}$$
(4)

and obtained by the analysis of the multiple-exponential decay profiles of fluorescence intensity, the following equation is inferred

$$\langle \tau \rangle = \frac{\tau_0 + \tau_\infty K \Phi [HPCD]_0^n}{1 + K \Phi [HPCD]_0^n}$$
(5)

When lifetime measurements are collected at the emission wavelength λ_1 , the parameter Φ from both Eqs. 2 and 5 has the same value.

From the steady-state fluorescence depolarization measurements (*L*-method) [51], the fluorescence anisotropy r can be quantified by

$$r = (I_{VV} - GI_{VH}) / (I_{VV} + 2GI_{VH})$$
(6)

 I_{xy} is the intensity of the emission that is measured when the excitation polarizer is in position x (V for vertical, H for horizontal), the emission polarizer is in position y, and the G factor (= I_{HV}/I_{HH}) corrects for any depolarization produced by the optical system. Under the assumption that the total anisotropy is the sum of contributions due to uncomplexed and complexed guests, the following equation can be derived

$$r = \frac{r_0 + r_\infty K \Phi[HPCD]}{1 + K \Phi[HPCD]}$$
(7)

It contains the same Φ parameter as the previous equations when anisotropies are collected at λ_1 and at the same excitation wavelength.

The dynamic quenching of a single excited species by a quencher (Q) follows the known Stern–Volmer equation [52]. Even for simple systems, Stern–Volmer plots of steady-state measurements are usually non-linear, nevertheless they can give information about the quencher to chromophore accessibility.

Molecular mechanics (MM)

Calculations were performed with Sybyl 6.9 and the Tripos Force Field [53, 54]. A relative permittivity $\varepsilon = 1$ was used in the vacuum and in the presence of water. Charges for 23DMN were obtained by MOPAC [55]. Geometry and charges for water, *HP*CDs were identical to those previously used [50]. Initial *HP*CDs, fully substituted at C2 of the glucopyranose unit (molar substitution=1.0), and 23DMN conformations were also taken from literature [49, 50]. Non-bonded cut-off distances were set at 8 Å, the simplex algorithm and the conjugate gradient (0.2 and 1.0 kcal/mol Å in the vacuo and water respectively) were used as optimization and termination methods [56, 57]. The molecular silverware algorithm was employed for water solvation of the systems [58]. Periodic boundary conditions were also employed. Interaction energy was obtained as the difference between the potential energy of the whole system and the sum of the potential energies of the isolated components in the same structure.

Results and discussion

Fluorescence measurements

Excitation spectra (at 385 nm) for 23DMN water solutions in the absence and in the presence of HPCDs showed bands at ~290 and ~335 nm. Emission spectra for 23DMN and 23DMN/HPCD water solutions recorded at λ_{ex} =282 nm, are depicted in Fig. 1. All spectra exhibit the two characteristic bands from naphthoate [20-22, 25-27, 39, 49] whose maxima slightly shifts to the blue upon increasing [HPCD]. Fluorescence intensity (area under emission spectrum) does not exhibit any special trend with [HPCD], as it increases or decreases depending on the host type. In addition, the spectra do not show any evidence of the inclusion of more than one 23DMN inside HPCDs. Nevertheless, the main feature takes place in the ratio of intensities of both bands, which is strongly dependent on the medium polarity. Such bands are centered around 362± 2 nm (λ_1) and 383±2 nm (λ_2). The parameter *R*, measured as $I_{\sim 383 \text{ nm}}/I_{\sim 362 \text{ nm}}$, substantially varies with [HPCD] and temperature. The top of Fig. 2 depicts the decreasing in Rwith [HPCD] at different temperatures for the three systems. The shape of these curves suggests that complexes of 23DMN with β HPCD are the most stable. Lines depicted in Fig. 2 come from the non-linear (top) and linear (bottom) adjustments of experimental data to Eqs. 2 and 3 respectively. The analysis agrees with a 1:1 stoichiometry and provides K, R_0 and R_∞ values by using $\Phi_{362 \text{ nm}}$ calculated at each temperature from the emission intensity at 362 nm in the absence of HPCD and at [HPCD] $\rightarrow \infty$. As Table 1 shows, $\Phi_{362} < 1$ for 23DMN/ α HPCD and 23DMN/ γ HPCD systems whereas Φ_{362} >1 for the 23DMN/ β HPCD one. In addition, Φ_{362} in the presence of $\beta HPCD$ ($\gamma HPCD$) seems to decrease (increase) with temperature. The value for 23DMN: α CD, however, is almost temperature independent. R_0 for 23DMN at each temperature, as expected, are very similar for all systems. R_{∞} , however, depends on the *HP*CD type and temperature. At 25 °C R_0 is 0.99, whereas R_{∞} are 0.79, 0.68 and 0.83 for α -, β - and γ *HP*CDs complexes. These values denote a



Fig. 1 Emission spectra for 23DMN/ α HPCD, 23DMN/ β HPCD and 23DMN/ γ HPCD systems at different [HPCD] concentrations at 25 °C. Concentrations range from 0 to 3.724×10^{-2} , 1.792×10^{-2} and 3.187×10^{-2} M for α -, β - and γ HPCD respectively. *Dashed curves* are for 23DMN in the absence of HPCDs

decreasing in the medium polarity surrounding 23DMN in the complex, which depends on the *HP*CD.

The average of *K* are around 165, 1,080 and 80 M⁻¹ for 23DMN complexes with α -, β - and $\gamma HPCDs$ at 25 °C. The value for 23DMN: $\beta HPCD$ at 25 °C is comparable to the one obtained for 23DMN: βCD (1,060 M⁻¹) [49]. The ones for 23DMN: $\alpha HPCD$ and 23DMN: $\gamma HPCD$ are somewhat larger than the one observed for 23DMN with the naturally occurring αCD (20 M⁻¹) and βCD (40 M⁻¹) [49].

Fluorescence intensity decay profiles ($\lambda_{exc}=282$ nm and $\lambda_{em}=362$ nm) for the 23DMN water solutions were monoexponentials, with a single fluorescence lifetime around 5 ns which hardly changes in the 5°-45 °C temperature range. Decays for 23DMN/*HP*CD water solutions at any temperature and [*HP*CD] were fitted to the sum of two exponentials. As a result, two lifetime components, a short one close to the value for the free guest (5 ns) and a longer one attributed to the complexed form and whose value



Fig. 2 (*Top*) Variation of *R* with [*HPCD*] and temperature. *Curves* are the result of the fitting by using Eq. 2; (*bottom*) double-reciprocal plots for 23DMN guest complexes at different temperatures by fitting

the data to Eq. 3. Symbols are a 5 °C (filled square), 15 °C (filled circle), 25 °C (filled triangle), 35 °C (inverted filled triangle) and 45 °C (filled diamond)

<i>T</i> (°C)	$K (\mathrm{M}^{-1})$	$\Phi_{362\ nm}$	R_0	R_∞
23DMN/αHPC	D			
5	213.1±15.3 (209.4±7.4)	0.73	0.994 ± 0.003	$0.780 {\pm} 0.004$
15	163.3±16.9 (168.5±7.4)	0.78	$0.989 {\pm} 0.004$	$0.782 {\pm} 0.006$
25	158.8±23.8 (169.5±9.9)	0.75	$0.988 {\pm} 0.005$	$0.786 {\pm} 0.008$
35	149.7±28.6 (150.1±10.7)	0.67	$0.980 {\pm} 0.006$	$0.789 {\pm} 0.011$
45	150.7±40.4 (152.7±14.6)	0.73	$0.975 {\pm} 0.007$	$0.831 {\pm} 0.011$
23DMN/βHPC	D			
5	1,676.6±51.9 (1,662.9±36.1)	1.45	0.992 ± 0.002	$0.693 {\pm} 0.001$
15	1,292.5±35.1 (1,273.9±27.8)	1.42	0.993 ± 0.002	$0.686 {\pm} 0.001$
25	1,100.7±38.5 (1,066.6±27.7)	1.42	$0.986 {\pm} 0.002$	$0.683 {\pm} 0.001$
35	1,035.8±29.7 (1,004.1±19.9)	1.30	$0.982 {\pm} 0.002$	$0.681 {\pm} 0.001$
45	841.1±26.9 (817.4±20.4)	1.25	$0.977 {\pm} 0.001$	$0.677 {\pm} 0.001$
23DMN/yHPCI)			
5	60.7 ±15.2 (-±-)	0.83	$1.028 {\pm} 0.008$	$0.834 {\pm} 0.016$
15	66.1±10.7 (44.8±4.9)	0.90	1.005 ± 0.003	$0.825 {\pm} 0.012$
25	95.8±11.2 (76.3±6.5)	0.95	$0.994 {\pm} 0.003$	$0.831 {\pm} 0.006$
35	112.3±8.7 (103.3±5.1)	0.99	$0.985 {\pm} 0.002$	$0.828 {\pm} 0.004$
45	115.4 ±9.3 (112.5±8.7)	1.07	$0.977 {\pm} 0.002$	0.822 ± 0.004

Table 1 Association constants K, obtained from the non-linear and linear adjustments (parentheses), R_0 and R_{∞} , as well as $\Phi_{362 \text{ nm}}$ for 23DMN/ αHP , 23DMN/ βHP CD and 23DMN/ γHP CD systems at different temperatures

depends on the CD type, were obtained. Figure 3 depicts the variation of the lifetime averages, $as\langle \tau \rangle / \tau_0$, with [HPCD] for one of the systems at several temperatures (top) and for the three systems at 25 °C (bottom). Curves show the result of the adjustments to a rearranged Eq. 5, by using Φ_{362} values obtained from steady-state measurements. K's obtained from this analysis are reasonably similar to the ones from steady state, *i.e.*, 180 ± 17 , $930\pm$ 35 and 115±24 M^{-1} for complexation with α -, β - and $\gamma HPCD$ respectively at 25 °C. Values of τ_{∞} , at the same temperature, are 6.7, 7.4 and 6.5 ns respectively. These values together with R_{∞} (0.79, 0.68, 0.83) for α -, β - and yHPCDs respectively at 25 °C can provide information about the microviscosity and polarity of the medium surrounding 23DMN when complexed. For this purpose the dependence of R and τ on the polarity (ε) and microviscosity (η) was obtained by performing emission spectra and time-resolved measurements for 23DMN dilute solutions of different hydroxypropylated solvents covering a wide range of ε and η at 25 °C [49]. Emission spectra showed the two typical bands and intensity decays are mono-exponential for any of the solvents used. R increases monotonically with ε according the following quadratic equation $R = 0.55 + 5.32 \times 10^{-4} \varepsilon + 6.14 \times 10^{-5} \varepsilon^2$, where-



Fig. 3 (*Top*) Changes in the average lifetime, $\langle \tau \rangle$ with [β*HP*CD] and temperature: Symbols are a 5 °C (*filled square*), 15 °C (*filled circle*), 25 °C (*filled triangle*), 35 °C (*inverted filled triangle*) and 45 °C (*filled diamond*); (*Bottom*) variation in $\langle \tau \rangle$ with [α*HP*CD] (*empty square*), [β*HP*CD] (*empty circle*) and [γ*HP*CD] (*empty triangle*) at 25 °C. *Curves* fit the experimental data to the Eq. 5 by assuming Φ_{362} nm values obtained from the emission spectra; (λ_{em} =362 nm, λ_{ex} = 282 nm). These fits provide values for *K* at 25 °C of 178.0±17.4; 929.8±34.5 and 115.1±24.0 M⁻¹ for 23DMN complexes with α-, βand γ*HP*CDs respectively

as it hardly changes with η at the same temperature. τ , however, decreases with ε and it increases almost linearly with η .

Effective dielectric constants (ε) for the media surrounding 23DMN in the complexes with α -, β - and γ HPCDs of 58, 41 and 63 were estimated from R_{∞} and the above equation. These values are quite similar to those obtained when 2MN complexes with α -, β - and γ HPCDs (51, 44 and 56 respectively) [21]. Something different occurs when 23DMN complexes with their counterparts α -, β - and γ CD [49]. The value for 23DMN/ α CD ($\epsilon \approx 70$) substantially differs from that for 23DMN/ α HPCD. 23DMN partially penetrates in the most hydrophobic aCD cavity and totally in the β - and γ CD ones [49]. However, 2MN, which is less bulky than 23DMN, seems to deeply penetrate inside the three α -, β - and γ *HP*CDs. Thus ε values, that which are very similar ($\varepsilon \approx 50$), correspond to the ones for the inner α -, β - and γ *HPCD* cavities. This contrasts with the natural CDs whose polarities are rather different (around 6, 50 and 70) [20, 21, 50]. As $\langle \tau \rangle / \tau_0$ increases with [HPCD] (and $\tau_{\infty} > \tau_0$), this increase may be attributed to the η increases and the ε decreases around the guest upon complexation. The largest τ_{∞} obtained for 23DMN/ β HPCD as compared to the other two complexes, may be a consequence of the highest microviscosity surrounding a guest that is deeply trapped inside a cavity with a relatively low ε (= 44).

 ΔH^0 and ΔS^0 were obtained from the average of *K* constants from steady-state measurements by using the linear van't Hoff representations depicted in Fig. 4. The results of the analysis were collected in Table 2 and compared with the thermodynamic parameters for 23DMN complexation with the unsubstituted CDs [49]. Inclusion processes for 23DMN with α - and $\beta HPCDs$ have $\Delta H^0 < 0$, but are slightly more favorable for the 23DMN/ $\beta HPCD$ complex. The 23DMN: γ CD is accompanied by an unfavorable positive enthalpy change. The signs of ΔH^0 match those obtained for complexation with their natural CDs, however absolute values are smaller as a consequence



Fig. 4 Van't Hoff representations for the formation of the 1:1 stoichiometry 23DMN: α HPCD (*empty square*), 23DMN: β HPCD (*empty circle*) and 23DMN: γ HPCD (*empty triangle*) complexes

 Table 2
 Thermodynamics parameters obtained from the average of the constants collected in Table 1 from non-linear and linear fits at different temperatures

Complex	$\Delta H^0 \ (\text{kJ mol}^{-1})$	$\Delta S^0 (\mathrm{JK}^{-1} \mathrm{mol}^{-1})$
α <i>HP</i> CD	-7.8 ± 2.5	+15.9±8.7
(aCD)	(-9.1 ± 5.2)	(-4.8 ± 15.0)
β <i>HP</i> CD	-12.0 ± 1.2	$+18.1\pm4.1$
(βCD)	(-15.7 ± 1.6)	$(+6.0\pm5.6)$
γ <i>HP</i> CD	$+14.1\pm3.3$	$+83.9\pm11.1$
(γCD)	(+35.2±7.5)	$(+145.9\pm25.2)$

In parentheses are the values obtained for the complexes of 23DMN with α -, β - and γ CDs from Ref. [49]

of the probably weaker guest/host interactions due to their larger cavity sizes.

The entropic terms accompanying the complexation are favourable for any of the 23DMN/HPCD complexes. Something similar occurred when 23DMN complexes with β - and γ CDs, but it contrasts with the result obtained when it complexes with α CD. This complexation was accompanied by a small negative entropy change [39, 49]. $\Delta S^0 < 0$ values are typically obtained for guests that partially penetrate inside the CD cavity and/or whose movement is hindered by strong intermolecular guest-CD interactions. A guest that penetrates into the cavity exhibits $\Delta S^0 > 0$. The entropy sign is normally the result of two opposite effects, the loss of the solvent order surrounding the guest or included inside the CD host, and the decrease in the degrees of freedom of the guest due to its association with the host upon complexation. Thermodynamic parameters together with the values of R_{∞} may agree with a good penetration of 23DMN inside any of the three hydroxypropylated hosts. The placement of the guest would depend on the HPCD size.

Figure 5 depicts changes in r with [HPCD] for 23DMN/ β *HP*CD at different temperatures and for the three systems at 25 °C. Curves are the results of the adjustments, which seem to fit reasonably, of experimental anisotropy data to Eq. 7 by using K and Φ values from steady-state. r increases as $[\beta HPCD]$ increases because of the larger fraction of the complexed form which has a smaller rotational diffusion rate than free 23DMN and it decreases with temperature. This is probably due to a decrease in the solvent viscosity with temperature instead of the decrease in the fraction of the complex, as this trend is also observed for 23DMN/ γ HPCD, whose complexation is accompanied by $\Delta H^0 > 0$. Values at [*HP*CD] $\rightarrow \infty$ (R_{∞}) at 25 °C obtained from the adjustments to Eq. 7, are 0.0062 ± 0.0002 , $0.0034\pm$ 0.0002 and 0.0097 \pm 0.0004 for complexes with α -, β - and γ *HP*CDs respectively. These values, which follow the same trend as when 23DMN complexes with unsubstituted CDs [49], are obviously larger than those obtained for the free 23DMN guest $(r_0 \approx 7 \times 10^{-4})$. By assuming that the whole complex, where 23DMN is located inside the cavity, moves as a whole, the larger value for R_{∞} ($\gamma HPCD$)> R_{∞} ($\beta HPCD$) may be related to the size of the complex. However, the relatively large R_{∞} ($\alpha HPCD$) value may agree with the fact that 23DMN would not totally be include inside $\alpha HPCD$ but tightly bonded to it, making the size of the complex relatively large.

Quencher measurements at 25 °C were performed on 23DMN and 23DMN/HPCD water solutions for a 0.75 fraction of the guest complexed by using a solution of diacetyl, (CH₃CO)₂ (1 M), as quencher. Emission intensity obviously decreases upon quencher addition, but R remains almost constant (the presence of the quencher does not substantially modify the structure of the complexes). Figure 6 shows Stern-Volmer representations for the 23DMN and 23DMN/HPCDs, which in the range of quencher concentration are non-linear. The results agree with the fact that the free 23DMN is much more accessible to the quencher than the complexed one. But they also reveal, in agreement with a not total penetration of 23DMN into the $\alpha HPCD$ cavity, that the accessibility of 23DMN in the complex with $\alpha HPCD$ is larger than with $\beta HPCD$, and this in turn is larger than when it complexes with $\gamma HPCD$.



Fig. 5 (*Top*) Variation of the fluorescence anisotropy, *r* with [β*HP*CD] at different temperatures: 5 °C (*filled square*), 15 °C (*filled circle*), 25 °C (*filled triangle*), 35 °C (*inverted filled triangle*) and 45 °C (*filled diamond*), measured at the maximum of the low energy band (λ_{em} = 362 nm, λ_{ex} =282 nm); (*bottom*) variation of *r* with [α*HP*CD] (*empty square*), [β*HP*CD] (*empty circle*) and [γ*HP*CD] (*empty triangle*) at 25 °C



Fig. 6 Stern–Volmer plots from steady-state (fluorescence intensity) at 25 °C by using diacetyl as quencher of 23DMN (*plus sign*) and 23DMN: α *HPCD (empty square*), 23DMN: β *HPCD (empty circle*) and 23DMN: γ *HPCD (empty triangle*) water solutions. Solutions were for a [*HPCD*] which provides a 0.75 fraction of the complexed guest

Circular dichroism measurements

Absorption spectrum for free 23DMN or in the presence of HPCDs exhibits three main bands placed at approximately 245, 282 and 320 nm which, according to the Platt's notation, were ascribed to the ${}^{1}B_{b}$, ${}^{1}L_{a}$ and ${}^{1}L_{b}$ bands respectively [59]. Transition moments are nearly parallel to the long naphthalene axis for ¹B_b and to the short one for ${}^{1}L_{a}$ and ${}^{1}L_{b}$ bands [60, 61]. Figure 7 depicts ICD spectra at 25 °C for isolated 23DMN and 23DMN/HPCD water solutions, upon selecting the intense ${}^{1}B_{h}$ naphthalene absorption band. All 23DMN/HPCD solutions were prepared for a 0.7 fraction of the complexed guest. Figure 7 also shows the absorption spectrum for 23DMN. The ICD spectrum for both 23DMN/BHPCD and 23DMN/yHPCD exhibits a positive band. This band is substantially more intense for the former one. A weak negative one is obtained for the 23DMN/ α HPCD solution and the ICD spectrum for the isolated 23DMN guest is obviously not observed. When focussing on the ${}^{1}L_{a}$ band (~275 nm) for 23DMN/ β HPCD at a higher 23DMN concentration, a faint negative band is obtained.

The ICD spectrum sign varies depending on the inclusion of the chromophore in the CD cavity and the orientation of its electronic transition moment relative to the CD n-fold rotational axis. Parallel (perpendicular) orientation gives a positive (negative) ICD band [34, 35, 62–65]. A large intensity is usually related to a better fit of the chromophore into de CD cavity. In addition, the sign of the ICD when the

chromophore is located partially outside the cavity becomes opposite to the inside one [66-68]. The positive ICD spectra for β - and γ HPCDs suggest that 23DMN guests are axially oriented, *i.e.*, long naphthalene axis nearly parallel to the *n*fold rotation CD axis. This is corroborated by the change of ICD sign for the ¹L_a band. In addition, the high intensity of the 23DMN/ β HPCD ¹B_b band suggests a better fit and deep penetration of 23DMN inside $\beta HPCD$, whose movement is probably quite hindered. The decreasing in intensity for the complex with γ *HP*CD probably comes from the increase in the cavity size resulting in a fit that is slightly worse [67, 69]. In contrast, the weak negative ICD band for the 23DMN/ α HPCD solution agrees, either with the fact that 23DMN do not penetrate totally inside the cavity, or that the 23DMN is equatorially oriented in the inner HPCD cavity, which is rather improbable [60].

A non-linear equation similar to Eqs. 2, 5 and 7, derived by assuming an ellipticity zero for free 23DMN and Φ =1, can also provide association constants. Experimental data adjust nicely to the non-linear equation showing K = 910 ± 50 M⁻¹ for 23DMN/βHPCD at 25 °C, which reasonably agrees with those obtained from different fluorescence techniques.



Fig. 7 Circular dichroism spectra of 23DMN and 23DMN in the presence of α -, β - and γ *HPCD* water solutions (prepared for a [*HPCD*] which provides a 0.7 fraction of complexed guest)



Fig. 8 Coordinate systems used to follow the (1:1) 23DMN complexation process with *HP*CDs by polar and non-polar guest-to-host approaching



Each HPCD was initially placed so that the center of mass of the glycosidic oxygen atoms (denoted by o in Fig. 8) was located at the origin of a coordinate system with its axis oriented as depicted [49, 50]. The 23DMN guest was then approached to the HPCD by the polar (P) and non-polar (NP) sides as shown in Fig. 8. The host-guest distance was measured as the oo' projection (d) on the y axis. The inclusion torsional angle (θ) was measured between the yz and oo'C9 (C9 from naphthalene group) planes. The oo'C9 angle (δ) also contributes to defining the orientation of 23DMN relative to the y axis. To obtain the most favorable approach of the guest, binding energies of all the optimized structures in the vacuum obtained by scanning δ and θ angles, in the $10-100^{\circ}$ and $0-55^{\circ}$ ranges respectively $(\Delta \delta = \Delta \theta = 10^{\circ})$ and the *d* distance from y = +10 (Å) to -6(Å) ($\Delta d=2$ Å), were calculated. Once this orientation was fixed 23DMN was approached to HPCD along the y coordinate from 20 (Å) through -20 (Å) at $\Delta d=0.5$ Å. Each structure generated was solvated and optimized.

Binding energies as a function of the d distance (along the y coordinate) between 23DMN and HPCD (polar and non-polar approaches) for three systems are depicted in Fig. 9. Calculations suggest that the complexation processes are favorable. Van der Waals interactions are mostly responsible for the complexation, while the electrostatics are insignificant at any host guest distance. The inclusion



Fig. 9 Binding energies as a function of guest-host distance d (in Å) for 23DMN complexation processes with α - (*top*), β - (*middle*) and γ CDs (*bottom* panel) by polar (*P*; *filled* circle) and non-polar (*NP*; *empty* circle) guest-to-host approaching

Fig. 10 MBE structures for the (1:1) 23DMN complexes with α *HPCD (1 and 2)*, β *HPCD (3) and \gammaHPCD (4) obtained from representation depicted in Fig. 9*

process of 23DMN into the α HPCD, due to its small cavity size, however, is accompanied by a large energetic gap. This does not occur with the complexation of the other two HPCDs, where the guest seems to favorably pass through the cavity, especially on the NP side. As a consequence the minima binding energy conformation structures for 23DMN: α HPCD are reached at relatively large v values (5.6 and 7.6 Å for both NP and P approaches respectively). For these structures a portion of 23DMN is outside the cavity and exposed to the water. Nevertheless, this portion is considerably smaller than it is when 23DMN complexes with its counterpart α CD [49]. However, 23DMN is located inside the β - or γ HPCD and perfectly shielded from the solvent (y=-1.6 and 0.3 Å respectively). These structures, depicted in Fig. 10, are in agreement with most of the experimental fluorescence and dichroism circular findings.

Conclusions

23DMN forms complexes of stoichiometry 1/1 with α -, β and $\gamma HPCDs$. The stability constants for the complexes with α - and $\gamma HPCDs$ are somewhat larger than those for complexes formed with their naturally occurring counterparts α - and γ CDs. Similar association constants for 23MN with $\beta HPCD$ or βCD complexes were obtained. Whereas the signs of ΔH^0 match those obtained with their unsubstituted counterparts, their absolute values are smaller, as a consequence of the probably weaker guest/ host interactions due to their larger cavity sizes. Favourable ΔS^0 for the three complexes could account for a good penetration of 23DMN inside de HPCDs cavities. Nevertheless, in case of $\alpha HPCD$, whereas tightly bonded to it, the inclusion may be not total. The latest statements will agree with the effective dielectric constant estimated when 23DMN complexes with $\alpha HPCD$, but also with R_{∞} values and the quenching and circular dichroism experiments. Experimental results are also in agreement with a total penetration of 23DMN inside β - and γ HPCD. MM calculations showed that 1:1 complexes of 23DMN with HPCDs are capable of forming and that the non-bonded van der Waals interactions are mainly responsible for complex stability. Minima binding structures for the complexes nicely corroborates experimental finding.

Acknowledgements This research was supported by the Comunidad de Madrid (CAM project: S-055/MAT/0227), CICYT (project CTQ2005-04710/BQU) and Consejería de Educación y Ciencia de la Junta de Castilla-La Mancha (grant to M.J.G-A). We are grateful to Dr. Krois from the University of Vienna for the time he dedicated to us (M.J.G-A). We wish to express our thanks to M.L. Heijnen for assistance with the preparation of the manuscript.

References

- Szejtli J, Osa T (1996) Comprehensive supramolecular chemistry, vol. 3. Cyclodextrins, Elsevier, Oxford
- 2. D'Souza VT, Lipkowitz KB (eds) (1998) Cyclodextrins. Chem Rev 98(5):1741–2076
- Harada A (2001) Cyclodextrin-based molecular machines. Acc Chem Res 34:456–464
- Nepogodiev SA, Stoddart JF (1998) Cyclodextrin-based catenanes and rotaxanes. Chem Rev 98(5):1959–1976
- Flamigni L (1993) Inclusion of fluorescein and halogenated derivatives in α-, β-, and γ-cyclodextrins: a steady-state and picosecond time-resolved study. J Phys Chem 97(38):9566–9572
- Fraiji EK Jr, Cregan TR, Werner TC (1994) Binding of 2acetylnaphthalene to cyclodextrins studied by fluorescence quenching. Appl Spectrosc 48(1):79–84
- Nakamura A, Sato S, Hamasaki K, Ueno A, Toda F (1995) Association of 1:1 inclusion complexes of cyclodextrins into homo- and heterodimers: a spectroscopic study using a TICTforming fluorescent probe as a guest compound. J Phys Chem 99 (27):10959–10959
- van Stam J, De Feyter S, De Schryver FC, Evans CH (1996) 2naphthol complexation by b-cyclodextrin: influence of added short linear alcohols. J Phys Chem 100(51):19959–19966
- Hamai S (1997) Inclusion of 2-chloronaphthalene by α-cyclodextrin and room-temperature phosphorescence of 2-chloronaphthalene in aqueous d-glucose solutions containing α-cyclodextrin. J Phys Chem B 101(9):1707–1712
- Madrid JM, Villafruela M, Serrano R, Mendicuti F (1999) Experimental thermodynamics and molecular mechanics calculations of inclusion complexes of 9-methyl anthracenoate and 1methyl pyrenoate with β-cyclodextrin. J Phys Chem B 103 (23):4847–4853
- Sadlej-Sosnowska N, Siemiarczuk A (2001) A time resolved and steady-state fluorescence quenching study on naproxen and its cyclodextrin complexes in water. J Photochem Photobiol A: Chem 138:35–40
- 12. Di Marino A, Mendicuti F (2004) Thermodynamics of complexation of dimethyl esters of tere-, iso-, and phthalic acids with α and β -cyclodextrins. Appl Spectrosc 58(7):823–830
- Shannigrahi M, Bagchi S (2005) Time resolved fluorescence study of ketocyanine dye-b cyclodextrin interactions in aqueous and non-aqueous media. Chem Phys Lett 403(1–3):55–61
- Turro NJ, Okubo T, Weed GC (1982) Enhancement of intramolecular excimer formation of 1,3-bichromophoric propanes via application of high pressure and via complexation with cyclodextrins. Protection from oxygen quenching. Photochem Photobiol 35(3):325–329
- Kano K, Takenoshita I, Ogawa T (1982) g-Cyclodextrin-enhanced excimer fluorescence of pyrene and effect of *n*-butyl alcohol. Chem Lett Chem Soc Jpn 3:321–324
- 16. Hamai S (1989) Pyrene excimer formation in γ -cyclodextrin solutions: association of 1:1 pyrene- γ -cyclodextrin inclusion compounds. J Phys Chem 93(17):6527–6529
- Pistolis G (1999) Dual excimer emission of *p*-terphenyl induced by g-cyclodextrin in aqueous solutions. Chem Phys Lett 304 (5,6):371–377
- Sainz-Rozas PR, Isasi JR, González-Gaitano G (2005) Spectral and photophysical properties of 2-dibenzofuranol and its inclusion complexes with cyclodextrins. J Photochem Photobiol A: Chem 173(3):319–327
- Catena GC, Bright FV (1989) Thermodynamic study on the effects of β-cyclodextrin inclusion with anilinonaphthalenesulfonates. Anal Chem 61(8):905–909

- Madrid JM, Mendicuti F, Mattice WL (1998) Inclusion complexes of 2-methylnaphthoate and γ-cyclodextrin: experimental thermodynamics and molecular mechanics calculations. J Phys Chem B 102(11):2037–2044
- Di Marino A, Mendicuti F (2002) Fluorescence of the complexes of 2-methylnaphthoate and 2-hydroxypropyl-α-, -β-, and -γcyclodextrins in aqueous solution. Appl Spectrosc 56(12):1579–1587
- 22. Pastor I, Di Marino A, Mendicuti F (2002) Thermodynamics and molecular mechanics studies on α- and β-cyclodextrins complexation and diethyl 2,6-naphthalenedicarboxylate guest in aqueous medium. J Phys Chem B 106(8):1995–2003
- Muñoz de la Peña A, Ndou TT, Zung JB, Warner IM (1991) Stoichiometry and formation constants of pyrene inclusion complexes with β- and γ-cyclodextrin. J Phys Chem 95(8):3330–3334
- 24. Will AY, Muñoz de la Peña A, Ndou TT, Warner IM (1993) Spectroscopic studies of the interaction of *tert*-butylamine and *n*propylamine with the β-cyclodextrin–pyrene complex. Appl Spectrosc 47(3):277–282
- 25. Madrid JM, Mendicuti F (1997) Thermodynamic parameters of the inclusion complexes of 2-methylnaphthoate and α and β -cyclodextrins. Appl Spectrosc 51(11):1621–1627
- 26. Cervero M, Di Marino A, Mendicuti F (2000) Inclusion complexes of dimethyl 2,6-naphthalenedicarboxylate with αand β-cyclodextrins in aqueous medium: thermodynamics and molecular mechanics studies. J Phys Chem B 104(7):1572–1580
- Pastor I, Di Marino A, Mendicuti F (2005) Complexes of dihexyl 2,6-naphthalenedicarboxylate with α- and β-cyclodextrins: fluorescence and molecular modeling. J Photochem Photobiol A: Chem 173(3):238–247
- Nelson G, Patonay G, Warner IM (1987) Fluorescence lifetime study of cyclodextrin complexes of substituted naphthalenes. Appl Spectrosc 41(7):1235–1238
- Ferreira JAB, Costa SMB (2005) Non-radiative decay in rhodamines: role of 1:1 and 1:2 molecular complexation with βcyclodextrin. J Photochem Photobiol A: Chem 173(3):309–318
- 30. Serna L, Di Marino A, Mendicuti F (2005) Inclusion complexes of a bichromophoric diester containing anthracene and naphthalene groups with alpha- and beta-cyclodextrins: thermodynamics and molecular mechanics. Spectrochim Acta Part A: Molec Biomolec Spectrosc 61(8):1945–1954
- Hossain MA, Mihara H, Ueno A (2003) Fluorescence resonance energy transfer in a novel cyclodextrin–peptide conjugate for detecting steroid molecules. Bioorg Med Chem Lett 13(24):4305–4308
- 32. Park JW, Lee SY, Kim SM (2005) Efficient inclusion complexation and intra-complex excitation energy transfer between aromatic group-modified β-cyclodextrins and a hemicyanine dye. J Photochem Photobiol A: Chem 173(3):271–278
- 33. Nigam S, Durocher G (1997) Inclusion complexes of some 3Hindoles with cyclodextrins studied through excited state dynamics and steady state absorption and fluorescence spectroscopy. J Photochem Photobiol A: Chem 103(1,2):143–152
- Krois D, Brinker UH (2006) Circular dichroism of cyclodextrin complexes. In: Dodziuk H (ed) Cyclodextrins and their complexes. Wiley-VCH, New York, pp 289–298 chapter 10.4
- 35. Murphy RS, Barros TC, Mayer B, Marconi G, Bohne C (2000) Photophysical and theoretical studies on the stereoselective complexation of naphthylethanols with β-cyclodextrin. Langmuir 16(23):8780–8788
- Lipkowitz KB (1998) Applications of computational chemistry to the study of cyclodextrins. Chem Rev 98(5):1829–1873
- Jursic BS, Zdravkovski Z, French AD (1996) Molecular modeling methodology of β-cyclodextrin inclusion complexes. J Mol Struct (Theochem) 366(1–2):113–117
- Salvatierra D, Jaime C, Virgili A, Sanchez-Ferrando F (1996) Determination of the inclusion geometry for the β-cyclodextrin/

benzoic acid complex by NMR and molecular modeling. J Org Chem 61(26):9578-9581

- 39. Madrid JM, Pozuelo J, Mendicuti F, Mattice WL (1997) Molecular mechanics study of the inclusion complexes of 2methyl naphthoate with α- and β-cyclodextrins. J Colloid Interface Sci 193(1):112–120
- Pozuelo J, Nakamura A, Mendicuti F (1999) Molecular mechanics study of the complexes of β-cyclodextrin with 4-(dimethylamino) benzonitrile and benzonitrile. J Incl Phenom Macroc Chem 35 (3):467–485
- 41. Cervello E, Mazzucchi F, Jaime C (2000) Molecular mechanics and molecular dynamics calculations of the β-cyclodextrin inclusion complexes with *m*-, and *p*-nitrophenyl alkanoates. J Mol Struct (Theochem) 530(1,2):155–163
- 42. Lino ACC, Takahata Y, Jaime C (2002) α- and β-cyclodextrin complexes with *n*-alkyl carboxylic acids and *n*-alkyl *p*-hydroxy benzoates. A molecular mechanics study of 1:1 and 1:2 associations. J Mol Struct (Theochem) 594(3):207–213
- 43. Pozuelo J, Mendicuti F, Mattice WL (1997) Inclusion complexes of chain molecules with cycloamyloses. 2. Molecular dynamics simulations of polyrotaxanes formed by Poly(ethylene glycol) and a-cyclodextrins. Macromolecules 30(12):3685–3690
- 44. Lipkowitz KB, Pearl G, Coner B, Peterson MA (1997) Explanation of where and how enantioselective binding takes place on permethylated β-cyclodextrin, a chiral stationary phase used in gas chromatography. J Am Chem Soc 119 (3):600–610
- 45. Köhler J, Hohla M, Söllner R, Eberle H-J (1998) Cyclohexadecanone derivative/γ-cyclodextrin complexes MD simulations and AMSOL calculations in vacuo and in aquo compared with experimental finding. Supramol Sci 5(1–2):101–116
- 46. Pozuelo J, Mendicuti F, Mattice WL (1998) Inclusion complexes of chain molecules with cycloamyloses. III. Molecular dynamics simulations of polyrotaxanes formed by polypropylene glycol and β-cyclodextrins. Polym J 30(6):479–484
- 47. Dodziuk H, Kozminski W, Lukin O, Sybilska D (2000) NMR manifestations and molecular dynamics modeling of chiral recognition of α-pinenes by α-cyclodextrin. J Mol Struct (Theochem) 525:205–212
- Grabuleda X, Ivanov P, Jaime C (2003) Computational studies on pseudorotaxanes by molecular dynamics and free energy perturbation simulations. J Org Chem 68(4):1539–1547
- 49. Alvariza C, Usero R, Mendicuti F (2007) Binding of dimethyl 2,3-naphthalenedicarboxylate with α -, β and γ -cyclodextrins in aqueous solution. Spectrochem Acta Part A 67(2):420–429
- Di Marino A, Mendicuti F (2007) Rationalizing some experimental facts on the complexation of 2-methyl naphthalenecarboxylate and hydroxypropyled cyclodextrins by molecular mechanics and molecular dynamics. J Incl Phenom Macrocyc Chem 58 (3–4):295–305
- Lakowicz JR (1999) Principles of fluorescence spectroscopy, 2nd edn. Kluwer, New York, p 298
- Valeur B (2002) Molecular fluorescence: principles and applications. Wiley-VCH, Weinheim, p 89
- 53. Sybyl 6.9; Tripos Associates; St. Louis, Missouri, USA
- Clark M, Cramer RC III, Van Opdenbosch N (1989) Validation of the general purpose Tripos 5.2 force field. J Comp Chem 10 (8):982–1012
- 55. MOPAC (AM1). Included in the Sybyl package
- Brunel Y, Faucher H, Gagnaire D, Rasat A (1975) Program of minimization of the empirical energy of a molecule by a simple method. Tetrahedron 31(8):1075–1091
- Press WH, Flannery BP, Teukolski SA, Vetterling WT (1988) Numerical recipes: the art of scientific computing. Cambridge University Press, Cambridge, p 312

- Blanco M (1991) Molecular silverware. I. General solutions to excluded volume constrained problems. J Comp Chem 12(2):237–247
- Platt JR (1949) Classification of spectra of cata-condensed hydrocarbons. J Chem Phys 17:484–495
- Kodaka M (1998) Application of a general rule to induced circular dichroism of naphthalene derivatives complexed with cyclodextrins. J Phys Chem 102(42):8101–8103
- Yorozu T, Hoshino M, Imamura M (1982) Fluorescence studies of pyrene inclusion complexes with α-, β-, and γ-cyclodextrins in aqueous solutions. Evidence for formation of pyrene dimer in γcyclodextrin cavity. J Phys Chem 86(22):4426–4429
- Harata K, Uedaira H (1975) Circular dichroism spectra of the βcyclodextrin complex with naphthalene derivatives. Bull Chem Soc Jpn 48(2):375–378
- Shimizu H, Kaito A, Hatano M (1979) Induced circular dichroism of b-cyclodextrin complexes with substituted benzenes. Bull Chem Soc Jpn 52(9):2678–2684

- 64. Shimizu H, Kaito A, Hatano M (1981) Induced circular dichroism of β-cyclodextrin complexes with *o*-, *m*-, and *p*-disubstituted benzenes. Bull Chem Soc Jpn 54(2):513–519
- 65. Tinoco I Jr (1962) Theoretical aspects of optical activity. II. Polymers. Adv Chem Phys 4:113–60
- 66. Kodaka M (1991) Sign of circular dichroism induced by βcyclodextrin. J Phys Chem 95(6):2110–2112
- Kodaka M (1993) A general rule for circular dichroism induced by a chiral macrocycle. J Am Chem Soc 115 (9):3702–3705
- Neckers DC, Volman DH, VonBünau G (1996) Advances in photochemistry. Wiley, New York, pp 5–35
- 69. Krois D, Brinker UH (1998) Induced circular dichroism and UV-vis absorption spectroscopy of cyclodextrin inclusion complexes: structural elucidation of supramolecular Azi-adamantane (Spiro[adamantane-2,3'-diazirine]). J Am Chem Soc 120(45): 11627–11632